**BioAim Scientific Inc** 

# Human BDNF EasyTest<sup>TM</sup> ELISA Kit

Cat.No: 1010003

**Instruction Manual** 

For research use only

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#### I. INTRODUCTION

Human mature BDNF is a 119 amino acid, 13 kDa, residue nonglycosylated polypeptide whose primary structure is conserved among all mammalian species examined. BDNF is 52% identical to NGF at the amino acid level. Cells known to express BDNF include fibroblasts, astrocytes, neurons, megakaryocytes/platelets, Schwann cells and smooth muscle cells. There are at least two receptors for BDNF, nerve Growth Factor Receptor (LNGFR), and TrkB (tropomyosin receptor kinase-B), that also binds NT-3 and NT-4/5.

LNGFR may serve as a retrograde transport molecule in neurons, promote Schwann cell migration near injury, and/or modulate TrkB activity in those cells that co-express both LNGFR and TrkB; While full TrkB activity is believed to require TrkB homodimerization, evidence suggests that full length TrkB and TrkC receptors may also form functional heterodimers in select cells where both receptors are co-expressed. These include cerebellar granule neurons and neurons of the hippocampal dentate nucleus. Among the cells known to express TrkB are motoneurons of the spinal cord, pyramidal cells of the hippocampus, almost all neurons in the developing brain, and thymocytes. During development, BDNF has been implicated in neuronal differentiation, maturation, survival and synapse formation. In the adult, one of its most promising roles centers on neuroprotection, possibly protecting forebrain neurons from ischemic attack and motor neurons from axotomy-induced death.

The BioAim Human BDNF EasyTest<sup>TM</sup> ELISA kit can quantitatively measure human BDNF in serum and plasma. It is a simple and rapid technology for the quantitation of antigen in a range of sample matrices. The whole process takes only 1.5 hours with high accuracy and precision. EasyTest<sup>TM</sup> ELISA is faster and easier to perform than standard format ELISA with less reagent handling and fewer pipetting steps.

#### II. REAGENTS

- 1. Human BDNF Microplate: 96 breakable wells (12strips x 8wells) coated with anti-human BDNF.
- 2. 20x Wash Buffer Concentrate: 1 Vial, 25 ml.
- 3. 5x Assay Diluent: 1 vial, 15 ml.
- 4. Standards: 10µl/ vial, 2 vials, recombinant human BDNF.
- 5. BioAim human BDNF Mix: 9µl/vial, 4 vials.
- 6. TBM Substrate solution: 1 Vial, 12 ml.
- 7. Stop Solution: 1 Vial, 8 ml of 0.2 M sulfuric acid.

## III. STORAGE

- 1. The kit can be stored for up to 6 months at 2° to 8°C from the date of shipment.
- 2. Standard can be stored at -20 °C or -80 °C. Use freshly prepared standard within 12hours (stored at 2~8°C).
- 3. Opened Microplate Wells or reagents may be store for up to 1 month at 2 to 8 °C. Return unused strip to the pouch containing desiccant pack, reseal along entire edge and keep in 2~8 °C.
- 4. Avoid repeated freeze-thaw cycles.

## IV. ADDITIONAL MATERIALS REQUIRED

- 1. Distilled or deionized water.
- 2. Precision pipettes, with disposable plastic tips.
- 3. Beakers, flasks, cylinders necessary for preparation of reagents.
- 4. Microplate washing device (multichannel pipette or automated microplate washer).
- 5. Microplate shaker.
- 6. Microplate reader capable of reading at 450 nm.

#### V. PRECAUTIONS

- 1. All reagents must be at room temperature (18 °C to 25 °C) before running assay.
- 2. Do not mix or substitute reagents with those from other lots or other sources.
- 3. Do not use kit reagents beyond expiration date on label.
- 4. Do not expose kit reagents to strong light during storage or incubation.
- 5. Use disposable pipette tips for each transfer to avoid microbial contamination or cross contamination of reagents.
- 6. Improper or insufficient washing at any stage of the procedure will result in either false positive or false negative results.
- 7. Avoid contact of stop solution with skin or eyes. If contact occurs, immediately flush area with copious amounts of water.
- 8. Do not use TMB substrate solution if it has begun to turn blue.
- 9. Do not expose bleach to work area during actual test procedure because of potential interference with enzyme activity.

#### VI. REAGENT PREPARATION

1. Bring all reagents and samples to room temperature (18~25°C) before use.

#### 2. Assay diluent

Dilute the concentrated assay diluent 1:5 with distilled water (e.g. 10ml plus 40ml).

#### 3. Wash Buffer

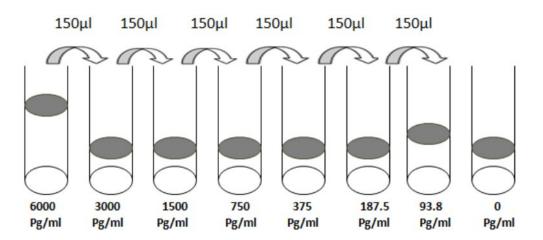
Dilute the concentrated wash buffer 1:20 with distilled water (e.g. 20ml plus 380ml).

#### 4. Sample

Levels of the target protein may vary among different specimens. Optimal dilution factors for each sample must be determined by the investigator. The dilution scheme is only suggestion: the recommended starting dilution for serum and plasma is 1: 5.

#### 5. Standard

- a. Briefly spin standard vial before use. Add 90 µl 1x Assay Diluent to prepare a 100ng/ml standard. Gently vortex to mix.
- b. Take 30 µl BDNF standard into a tube; then add 470 µl 1x Assay Diluent to prepare a 6000 pg/ml stock standard solution.
- c. Add 150 μl 1x Assay Diluent to 7 tubes. Label as 3000pg/ml, 1500pg/ml, 750pg/ml, 375pg/ml, 187.5pg/ml, 93.8pg/ml and the last tube with 1x assay diluent is the blank as 0pg/ml.
- d. Perform serial dilutions by adding 150  $\mu$ l of each standard to the next tube and vortexing between each transfer (see figure below).



#### 6. BioAim human BDNF Mix

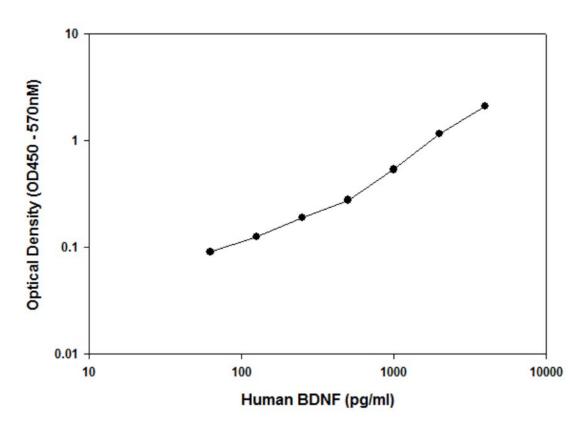
Within 15 minutes prior to use, briefly spin the vial. Add 1490  $\mu$ l of 1x Assay diluent to the vial and mix by pipetting. A vial mix can be used for around 30 wells.

#### VII. ASSAY PROCEDURE

- 1. All reagents must be brought to room temperature (18-25°C) prior to use. Place the required number of microwells in the holder. It is recommended that all samples, standards, and blanks be run in duplicate.
- 2. Add 50  $\mu$ l of 1x Assay Diluent into the blank wells.
- 3. Add 50 µl of each standard (*see reagent preparation step 5*) and samples into the designated wells. Gently shake/tap the plate for 5 seconds to mix.
- 4. Add 50  $\mu$ l of BioAim BDNF Mix into all wells, including the blank wells.
- 5. Cover wells with plate sealer and incubate at room temperature (18~25°C) for 1 hour with gentle shaking.
- 6. Decant or aspirate contents of wells. Wash wells by filling with at least  $300 \mu$ l/well prepared wash buffer followed by decanting/aspirating. Soak wells in wash buffer for 30 seconds to 1 minute for each wash. Repeat wash 4 times for a total of 5 washes. After the last wash, blot plate on absorbent paper to remove residual buffer. Thorough washing at this step is very important, complete removal of liquid is required for proper performance.
- 7. Pipette 100  $\mu$ l of TMB Substrate Solution to each well. Incubate plate for 15 minutes at room temperature in the dark with gentle shaking.
- 8. Add 50 µl of stop solution to each well.
- 9. Read absorbance at 450nm within 30 minutes of stopping reaction. If wavelength correction is available, subtract the optical density readings at 570nm from readings at 450nm.

#### VIII. CALCULATION OF RESULTS

- 1. Calculate the average absorbance values for each set of duplicate standards, samples and controls. Subtract the average zero standard optical density.
- 2. Create a standard curve by plotting the mean absorbance for each standard concentration on the ordinate against the BDNF concentration on log-log graph paper or using Sigma plot software. Draw a best fit curve through the points of the graph.
- 3. To determine the concentration of circulating BDNF for each sample, first find the mean absorbance value on the ordinate and extend a horizontal line to the standard curve. At the point of intersection, extend a vertical line to the abscissa and read the corresponding BDNF concentration.
- 4. A representative standard curve is shown below. This standard curve is for demonstration only. A standard curve must be run with each assay by operator.



#### **IX. PERFORMANCE**

#### A. Sensitivity

The minimum detectable dose of BDNF was determined to be 1pg/ml. This is defined as two standard deviations above the mean optical density of 20 replicates of the zero standards.

#### B. Recovery

Recovery was determined by spiking various levels of Human BDNF into the diluted sample types listed below. Mean recoveries are as follows:

| Sample Type | Average % recovery | Range % |
|-------------|--------------------|---------|
| Serum       | 122                | 102-135 |
| Plasma      | 115                | 109-123 |

C. Linearity

| Sample | Dilution | % of expected |
|--------|----------|---------------|
|        | 1:2      | 114           |
| Seum   | 1:4      | 113           |
|        | 1:8      | 132           |
|        | 1:2      | 96            |
| Plasma | 1:4      | 101           |
|        | 1:8      | 110           |

#### D. Specificity

No cross-reactivity was identified with the following cytokines: Adiponectin, Angiopoietin-1, Angiopoietin-2, IL-1 beta, IL-2, IL-3, IL-4, IL-5, IL-7, IL-8, IL-9, IP-10, G-CSF, GM-CSF, IFN-gamma, Leptin, MCP-1, PDGF, RANTES, SCF, TGF-beta, TIMP-2, TNF-alpha, TNF-beta, and VEGF.

E. Reproducibility Intra-Assay CV%: <10% Inter-Assay CV%: <15%

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## XI. Troubleshooting

| Problem               | Cause  | Solution   |
|-----------------------|--|--|
| 1.Poor standard curve | <ol> <li>Inaccurate pipetting</li> <li>Improper standard<br/>dilution</li> </ol>                                   | <ol> <li>check pipettes;</li> <li>Ensure briefly spin the vial of standard, take the right amount to dilution.</li> </ol>                                    |
| 2. Low signal         | <ol> <li>Too brief incubation<br/>time</li> <li>Inadequate reagent<br/>volumes or improper<br/>dilution</li> </ol> | <ol> <li>ensure adequate<br/>incubation time;</li> <li>Check pipettes and<br/>ensure corrected<br/>preparation.</li> </ol>                                   |
| 3. Large CV           | Inaccurate pipetting   | <ol> <li>Check pipettes;</li> <li>Accurately perform<br/>each step.</li> </ol>   |
| 4.High background     | <ol> <li>Plate is insufficiently<br/>washed;</li> <li>Wash buffer<br/>contamination</li> </ol>                     | <ul> <li>1.Follow the manual correctly; if using a plate washer, check that all ports are working functionally;</li> <li>2. Prepare fresh buffer.</li> </ul> |
| 5. Low sensitivity    | <ul><li>1.ELISA kit improper<br/>storage</li><li>2. Stop solution</li></ul>  | <ol> <li>Follow the manual to<br/>store each component<br/>correctly;</li> <li>Add enough stop<br/>solution to each well.</li> </ol>                         |

## **Bioaim Scientific Inc**

Unit 6, 27 Casebridge Court Scarborough, ON, M1B 4Y4 Canada Tel: 416-286-6868 www.bioaimscientific.com