

Synaptotagmin 1 luminal domain

Cat.No. 105 311CpH; Monoclonal mouse antibody, 100 µg purified IgG (lyophilized)

Data Sheet

Reconstitution/Storage	100 µg purified IgG, lyophilized, fluorescence-labeled with CypHer5E. Rabbit serum albumin was added for stabilization. For reconstitution add 100 µl H ₂ O to get a 1mg/ml solution in PBS. Either add 1:1 (v/v) glycerol, then aliquot and store at -20°C until use, or store aliquots at -80°C without additives. Reconstitute immediately upon receipt! Avoid bright light when working with the antibody to minimize photo bleaching of the fluorescent dye.
Applications	WB: N/A IP: N/A ICC: 1 : 50 up to 1 : 300 IHC: not tested yet IHC-P/FFPE: not tested yet EM: N/A ELISA: N/A FACS: not tested yet
Label	CypHer5E
Clone	604.2
Subtype	IgG1 (κ light chain)
Immunogen	Synthetic peptide corresponding to AA 1 to 12 from rat Synaptotagmin1 (UniProt Id: P21707)
Epitop	Epitop: AA 1 to 12 from rat Synaptotagmin1 (UniProt Id: P21707)
Reactivity	Reacts with: rat (P21707). No signal: mouse (P46096), zebrafish. Other species not tested yet.
Specificity	Specific for rat synaptotagmin 1, no cross-reactivity to other synaptotagmins.
Remarks	This antibody is intended to be used for direct labeling of recycling synapses in primary neuronal cultures. The pH sensitive dye regains its fluorescence after the reacidification of the synaptic vesicle lumen.

TO BE USED IN VITRO / FOR RESEARCH ONLY
NOT TOXIC, NOT HAZARDOUS, NOT INFECTIOUS, NOT CONTAGIOUS

Synaptotagmin 1 also known as **p65**, is an integral membrane glycoprotein of neuronal synaptic vesicles and secretory granules of neuroendocrine cells that is widely (but not ubiquitously) expressed in the central and peripheral nervous system. It has a variable N-terminal domain that is exposed to the lumen of the vesicle and a conserved cytoplasmic tail that contains two Ca²⁺-binding C2-domains. Ca²⁺-binding to synaptotagmin triggers exocytosis of synaptic vesicles, thus linking Ca²⁺-influx during depolarization to neurotransmitter

release.

Luminal antibodies were used in living neurons to label synaptic vesicles from the outside via endocytotic uptake.

Selected References SYSY Antibodies

Key physiological parameters dictate triggering of activity-dependent bulk endocytosis in hippocampal synapses. Wenzel EM, Morton A, Ebert K, Welzel O, Kornhuber J, Cousin MA, Groemer TW PLoS one (2012) 7(6): e38188. **UPTAKE**

Synapse clusters are preferentially formed by synapses with large recycling pool sizes. Welzel O, Tischbirek CH, Jung J, Kohler EM, Svetlitchny A, Henkel AW, Kornhuber J, Groemer TW PLoS one (2010) 5(10): e13514. **ICC**

Newly produced synaptic vesicle proteins are preferentially used in synaptic transmission. Truckenbrodt S, Viplav A, Jähne S, Vogts A, Denker A, Wildhagen H, Fornasiero EF, Rizzoli SO The EMBO journal (2018) : . **UPTAKE; tested species: rat**

Riluzole attenuates the efficacy of glutamatergic transmission by interfering with the size of the readily releasable neurotransmitter pool. Lazarevic V, Yang Y, Ivanova D, Fejtova A, Svenningsson P Neuropharmacology (2018) : . **ICC; tested species: rat**

Regulated Dynamic Trafficking of Neurexins Inside and Outside of Synaptic Terminals. Neupert C, Schneider R, Klatt O, Reissner C, Repetto D, Biermann B, Niesmann K, Missler M, Heine M The Journal of neuroscience : the official journal of the Society for Neuroscience (2015) 35(40): 13629-47. **ICC**

Dynamic properties of the alkaline vesicle population at hippocampal synapses. Röther M, Brauner JM, Ebert K, Welzel O, Jung J, Bauereiss A, Kornhuber J, Groemer TW PLoS one (2014) 9(7): e102723. **ICC; tested species: rat**

Blocking endocytosis enhances short-term synaptic depression under conditions of normal availability of vesicles. Hua Y, Woehler A, Kahms M, Haucke V, Neher E, Klingauf J Neuron (2013) 80(2): 343-9. **UPTAKE; tested species: rat**

The pH probe CypHerTM5E is effectively quenched by FM dyes. Welzel O, Loy K, Tischbirek CH, Tabor A, Gmeiner P, Kornhuber J, Groemer TW Journal of fluorescence (2013) 23(3): 487-94. **UPTAKE; tested species: rat**

Independent vesicle pools underlie different modes of release during neuronal development. Andrae LC, Fredj NB, Burrone J The Journal of neuroscience : the official journal of the Society for Neuroscience (2012) 32(5): 1867-74. **UPTAKE**

Use-dependent inhibition of synaptic transmission by the secretion of intravesicularly accumulated antipsychotic drugs. Tischbirek CH, Wenzel EM, Zheng F, Huth T, Amato D, Trapp S, Denker A, Welzel O, Lueke K, Svetlitchny A, Rauh M, et al. Neuron (2012) 74(5): 830-44. **UPTAKE**

A readily retrievable pool of synaptic vesicles. Hua Y, Sinha R, Thiel CS, Schmidt R, Hüve J, Martens H, Hell SW, Egner A, Klingauf J Nature neuroscience (2011) 14(7): 833-9. **UPTAKE**

Amyloid precursor protein is trafficked and secreted via synaptic vesicles. Groemer TW, Thiel CS, Holt M, Riedel D, Hua Y, Hüve J, Wilhelm BG, Klingauf J PLoS one (2011) 6(4): e18754. **UPTAKE**

Systematic heterogeneity of fractional vesicle pool sizes and release rates of hippocampal synapses. Welzel O, Henkel AW, Stroebel AM, Jung J, Tischbirek CH, Ebert K, Kornhuber J, Rizzoli SO, Groemer TW Biophysical journal (2011) 100(3): 593-601. **UPTAKE**

A common origin of synaptic vesicles undergoing evoked and spontaneous fusion. Hua Y, Sinha R, Martineau M, Kahms M, Klingauf J Nature neuroscience (2010) 13(12): 1451-3. **UPTAKE**

Selected General References

RAB3 and synaptotagmin: the yin and yang of synaptic membrane fusion. Geppert M, Südhof TC Annual review of neuroscience (1998) 21: 75-95.

The synaptic vesicle cycle: a cascade of protein-protein interactions. Südhof TC Nature (1995) 375(6533): 645-53.